Proceedings of the Iowa Academy of Science

Volume 66 | Annual Issue

Article 20

1959

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Wallace H. Orgell Iowa State University

Kunda A. Vaidya Iowa State University

Eugene W. Hamilton *Iowa State University*

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Recommended Citation

Orgell, Wallace H.; Vaidya, Kunda A.; and Hamilton, Eugene W. (1959) "A Preliminary Survey of Some Midwestern Plants for Substances Inhibiting Human Plasma Cholinesterase in Vitro," *Proceedings of the Iowa Academy of Science*, *66(1)*, 149-154.

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A Preliminary Survey of Some Midwestern Plants for Substances Inhibiting Human Plasma Cholinesterase *in Vitro*¹

By WALLACE H. ORGELL, KUNDA A. VAIDYA and

EUGENE W. HAMILTON

Abstract. Substances which inhibit the action of human plasma cholinesterase in vitro have been found in aqueous extracts of various members of the plant family Solanaceae. A preliminary survey of some weedy and cultivated midwestern representatives of this family revealed eleven active species in six genera: Datura, Lycopersicon, Nicotiana, Petunia, Physalis, and Solanum. A member each of the genera Capsicum and Lycium were inactive. Inhibitory substances were not found in thirty-one species representing nineteen plant families other than Solanaceae.

Substances which inhibit the action of the enzyme cholinesterase in vitro have been found in aqueous extracts of potato (Solanum tuberosum L.) foliage (Orgell, 1958a). Substances with a similar effect were subsequently found in tomato (Lycopersicon esculentum Mill.) and eggplant (Solanum melongena L.) extracts. These observations resulted in a preliminary examination of representatives of several plant families available in the vicinity of Ames, Iowa.

MATERIALS AND METHODS

The methods of collection, extraction, and assay were similar to those described previously (Orgell, 1958b). Plants were collected from the field and extracted within two hours after harvest. Ten grams of fresh tissue were homogenized in a Waring Blendor with 40 ml. of distilled water for 3 minutes at room temperature. The homogenate was filtered through a cotton plug, and the volume of filtrate measured. The filtrate was brought to pH 7.35 with 0.1 N sodium hydroxide, and diluted to 1.1 times its initial volume. Where semi-quantitative comparisons of many tissues were desired, 10 ml. aliquots of each filtrate (extract) were removed for assay. When quantitative determinations on single tissues were made, logarithmic series of five aliquots ranging from 0.01 to 3.16 ml., were taken for assay.

The various aliquots of aqueous plant extracts were each mixed with 5 ml. of standardized human blood plasma and made up to a total volume of 50 ml. with distilled water. The solutions were

¹Journal Paper No. J-3629 of the Iowa Agricultural and Home Economics Experiment Station, Ames, Iowa. Project No. 1351. This investigation was supported in part by Research Grant RG-4066 from the Division of Research Grants, U. S. Public Health Service.

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Table 1

Inhibition of Plasma	Cholinesterase	by	Aqueous	Extracts	of	Various
Plant Tissues						

Scientific name	Common name	Tissue	% I
Amaranthaceae Amaranthus retroflexus L.	Pigweed	Leaves Roots	0
Asclepiadaceae	Milkweed	Leaves	0
Chenopodiaceae	MIRWeed	Deaves	0
Chenopodium album L.	Lamb's quarters	Leaves	0
Compositae	D 1 1	T	0
Arctium minus Benth.	Burdock	Leaves	flowers 0
Taraxacum officinale Weber	Dandelion	Tops	
Cruciferae	Dundenon	1000	0
Brassica oleracea L.			
var. botrytis	Broccoli	Head	2
Brassica oleracea L.	Cabbaga	Head	0
var. capitata	Cabbage	Roots	2
Brassica napobrassica Mill.	Rutabaga	Roots	Ō
Cucurbitaceae			
Cucurbita pepo L.	Saurah	Logues	0
var. metopepo	Squasn	Fruits	0
		Roots	0
Cucurbita pepo L.	Conn. field		
	pumpkin	Fruit	0
		Roots	2
Convolvulaceae		10005	0
Convolvulus arvensis L.	Morning glory	Leaves	0
Equisetaceae	Hangatail	Shoots	0
Equiseium arvense L.	norsetan	Shoots	0
Ricinus communis L.	Castor bean	Leaves	0
Gramineae			
Zea mays L.	Corn	Foliage	0
Iridaceae	Gladiolus	Corm	0
Iris sp.	Wild iris	Leaves	Ő
Labiatae		_	
Nepeta cataria L.	Catnip	Leaves	0
Leguminosae Bhasaolus limensis Macf	Lima bean	Foliage	0
F naseotus umensis maci.	Enna Scan	Roots	Õ
		Pods	0
Trifolium repens L.	White Dutch clover	Tops	46
Medicago sativa L.	Alfalfa	rops	0
Allium sh	Iapanese onion	Leaves	0
Malvaceae	5.11		
Hibiscus trionum L.	Flower-of-an-hour	Leaves	0
Abutilon theophrasti Medic.	Velvet weed	Leaves	0
Polygonum bennsylvanicum L.	Smartweed	Leaves	0
Rumex crispus L.	Sourdock	Leaves	0
Portulacaceae	Duulana	Laguag	0
Portulaca oleracea L.	Pursiane	Roots	0

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Scientific name	Common name	Tissue	% I
Rosaceae			
Malus sylvestris Mill.	Jonathan apple	Fruits	0
Fragaria chiloensis	a. 1		
var. ananassa Duchesne	Strawberry	Fruits	1
Solanaceae	D ((])		
Solanum rostratum Dunal	Buffalo bur	Stem, seed pods	8
Solanum tuberosum L.	White Russet	uly leaves	0
	potato	Tuber flesh	40
	•	Tuber peels	58
		Peels $+$ flesh	51
		Peels + eyes	53
		Koots Leaves	41
		Flowers	50
		Berries	53
	Irish cobbler	Tuber peels + flesh	56
	Cherokee potato	Tuber peels + flesh	49
	Red Lasoda potato	Tuber peels $+$ flesh	54
	Red Pontiac potato	Tuber peels + flesh	40
Solanum melongena L. Potunia hybrida Vilm	Egg plant	r_{ruit}	19
retunta nyortaa viini.	retuilla	Flowers	55
Nicotiana tabacum L.	Tobacco	Leaves	5
Nicotiana glauca		Leaves	43
Nicotiana glutinosa		Leaves	31
Nicotiana bigloveii	T ' 1	Leaves	54
Datura stramonium L.	Jimson weed	Leaves Seed pode	20
		Roots	0
Physalis sp.	Ground cherry	Leaves + flowers	9
	,	Leaves	0
		Berries	35
Solanum carolinense L.	Horse nettle	Leaves + flowers	3
		Leaves	- 0
Capsicum trutascans I	Denner	Leaves	44
Capsicum fratescens L.	repper	Fruits	ŏ
		Roots	Ō
Solanum americanum Mill.	Nightshade	Leaves	30
		Berries	27
Solanum dulcamara L.	Nightshade	Lonvor	20
	Dittersweet	Berries	30
Lucium halimitolium Mill	Matrimony vine	Leaves	ŏ
		Berries	0
Lycopersicon esculentum Mill.	Tomato	Ripe fruit	0
		Green fruit	0
		Leaves	9 21
Umballiferae		ROOLS	41
Daucus carota L	Carrot	Leaves	0
		Roots	0
Xanthaceae		_	
Xanthium pennsylvanicum L.	Cocklebur	Leaves	0

Table I (Continued)

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stirred for 40 minutes at room temperature, during which time inhibition of the enzyme occurred. One ml. aliquots of the partially inhibited enzyme solutions were then removed for assay by the electrometric method of Curry (1956). Results were expressed either as the per cent inhibition resulting from 10 ml. of plant extract, or as the quantity of extract giving 50 per cent inhibition of the cholinesterase present in 5 ml. of plasma (I-50). Individual per cent inhibition values were corrected for an effect of the buffer capacity of the plant extract upon the electrometric assay, whereas the I-50 values were not corrected and are relative only to each other.

RESULTS

Assay of 44 plant species representing 21 families revealed cholinesterase-inhibitory substances in individuals from two of the families, the Leguminosae and the Solanaceae (Table 1). Cholinesterase inhibitors have been previously reported in two legumes. The calabar bean (*Physostigma venenosum* Balf.) is native to the west coast of Africa and is the source of the pharmacologically active alkaloid physostigmine (eserine). Ladino clover (*Trifolium repens* L.) contains an inhibitor which is reported to be soluble in chloroform (Heath, 1953). We found a similar inhibitor in extracts of White Dutch clover.

The eleven active species of the Solanaceae occurred in six genera: Datura, Lycopersicon, Nicotiana, Petunia, Physalis, and Solanum. All six species representing the genus Solanum were active. Garden pepper (Capsicum frutescens L.) and matrimony vine (Lycium halimifolium Mill.) were inactive. There appears to be no correlation between the poisonous character of some of these plants and their content of cholinesterase-inhibitor.

Table 2

Relative Concentration of Cholinesterase-Inhibitory Substances Present In Extracts of Solanaceous Plants

Plant	Variety or Cross	Tissue	I-50(ml)
Petunia		foliage	0.03
Potato	AI-5561-8	foliage	0.08
Potato	I-5569	foliage	0.16
Potato	AI-5561-13	foliage	0.23
Potato	Irish Cobbler	tuber sprouts	0.53
Potato	Irish Cobbler	foliage	0.56
Potato	Irish Cobbler	tuber peel	0.63
Potato	Plymouth	foliage	0.60
Potato	Huron	foliage	0.69
Horse nettle		berries	2.8
Common nightshade		berries	3.3
Ground cherry		berries	7.0
Potato	White Russet	tuber flesh	32.1

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The most potent source of inhibitory substances examined was the foliage of petunia (Table 2). As little as six milligrams of fresh petunia leaf resulted in 50 per cent inhibition of the cholinesterase present in 5 ml. of plasma. Foliage of certain experimental selections of potato were next in order of activity. Leaves, tuber skins, and tuber sprouts of commercial potato varieties contained lesser concentrations of inhibitor. Berries of horse nettle, common nightshade, and ground cherry were quite low in activity. Inhibitory substances of the potato plant were present in highest concentration in the foliage, flowers, berries, tuber sprouts, and skins. Lower levels were found in the stems and roots. The innermost flesh of potato tubers was practically devoid of inhibitors.

The distribution of cholinesterase-inhibitory substances within the plant varied from species to species within the Solanaceae. Inhibitory material was found primarily in the roots of garden tomato, with only low concentrations in the leaves and no activity in the fruits. The leaves of nightshade bittersweet contained inhibitor whereas none was present in the berries. The converse was true for the ground cherry. Leaves taken from different parts of the same field grown potato plant contained nearly equivalent levels of inhibitory substances.

Comparisons of the level of inhibitor in the foliage of various commercial varieties and in selections from a breeding program revealed considerable differences. In general, the common commercial varieties contained the lowest, and certain of the experimental crosses contained the highest levels of inhibitor. The AI-5561 series, which are partially derived from *Solanum chacoense*, are particularly rich in these substances.

Little is known about the effects of various environmental and climatic factors upon the concentration of inhibitory substances in potatoes. Plants grown in the greenhouse during the winter appear to have only one-half to one-fourth as much inhibitory material as those grown in the spring and summer in the field (Table 3). One experiment has indicated that the "greening" of potato tubers under artificial light results in an increased level of inhibitor in the tuber.

Potato	Field	Greenhouse	Field
Selection	(I-50)	(I-50)	Greenhouse
Plymouth	0.60	2.17	3.6
AI-5561-4	0.09	0.29	3.2
White Rose	1.16	2.58	2.2

 Table 3

 Relative Concentrations of Inhibitory Substances Present in Foliage of Potato

Selections Grown Under Field and Greenhouse Conditions

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DISCUSSION

Cholinesterase-inhibitory substances have been reported by others to be present in various plants, including raspberry (Beckett, 1954), boxwood (Vincent, 1945), and gelsemium (Vincent, 1951). Hence the presence of cholinesterase-inhibitory compounds is not unusual among higher plants. As yet, insufficient information is available to allow generalization with respect to the taxonomic distribution of these substances within the various plant families. The physical and chemical properties of these inhibitors suggest that they are alkaloids or alkaloidal glycosides, and that the substances from different families are also different chemically. The physiological and ecological role of these natural enzyme-inhibitors is at present obscure. It is perhaps possible that they play a part in the interactions of these plants with their predators and parasites.

Literature Cited

- Beckett, A. H., F. W. Belthle, and K. R. Fell. 1954. The active constituents of raspberry leaves, a preliminary investigation. Jour. Pharmacy and Pharmacology 6: 785-796.
- Curry, A. N. 1956. Procedure for the determination of residues of cholinesterase-inhibiting insecticides and their metabolites in plants. American Cyanamid Co., Analytical Research Lab., Stamford, Conn.
- Heath, D. F. and P. O. Park. 1953. An irreversible cholinesterase inhibitor in white clover. Nature (Lond.) 172: 206.
- Orgell, Wallace H., Kunda A. Vaidya, and P. A. Dahm. 1958a. Cholinesterase inhibition *in vitro* by extracts of potato. Proc. Iowa Acad. Science 65: 160-162.
- Orgell, Wallace H., Kunda A. Vaidya, and P. A. Dahm. 1958b. Inhibition of human plasma cholinesterase *in vitro* by extracts of solanaceous plants. Science 128: 1136-1137.
- Vincent, D. and T. Mathou. 1945. Inhibiting action of boxwood Buxus sempervirens and Buxus balearica on the cholinesterase of serum. Compt. rend. 220: 474-476.
- Vincent, D. and R. Lagreu. 1951. Action of the alkaloids of *Gelsemium* sempervirens on cholinesterases. Compt. rend. soc. biol. 145: 348-50.

Department of Zoology and Entomology Iowa State University Ames, Iowa